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# Structural basis for inhibition of the RNA-dependent RNA polymerase from SARS-CoV-2 by remdesivir

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The pandemic of Corona Virus Disease 2019 (COVID-19) caused by SARS-CoV-2 has become a global crisis. The replication of SARS-CoV-2 requires the viral RNA-dependent RNA polymerase (RdRp), a target of the antiviral drug, Remdesivir. Here we report the cryo-EM structure of the SARS-CoV-2 RdRp either in the apo form at 2.8 Å resolution or in complex with a 50-base template-primer RNA and Remdesivir at 2.5 Å resolution. The complex structure reveals that the partial double-stranded RNA template is inserted into the central channel of the RdRp where Remdesivir is covalently incorporated into the primer strand at the first replicated base pair and terminates chain elongation. Our structures provide critical insights into the mechanism of viral RNA replication and a rational template for drug design to combat the viral infection.

The COVID-19 pandemic caused by the virus SARS-CoV-2 has become a humanitarian crisis, with over 1.5 million infections and 87,000 death as reported on April 8 of 2020 (1, 2), which has increased rapidly to over 2.99 million infections and 207,000 death on April 27 of 2020 (2). SARS-CoV-2 is closely related to severe acute respiratory syndrome coronavirus (SARS-CoV) and several members from the beta coronavirus family, including bat and pangolin coronaviruses (3–5). Possibly due to stronger binding affinity of the virus spike protein for the host receptor (6–10), SARS-CoV-2 has much higher incidence of human to human transmission, resulting in infection worldwide.

SARS-CoV-2 is a positive strand RNA virus. Its replication is mediated by a multi-subunit replication/transcription complex of viral non-structural proteins (nsp) (11). The core component of this complex is the catalytic subunit (nsp12) of an RNA-dependent RNA polymerase (RdRp) (12, 13). By itself, nsp12 has little activity and its functions require accessory factors including nsp7 and nsp8 (14, 15), which increase RdRp template binding and processivity. RdRp is also proposed to be the target of a class of antiviral drugs that are nucleotide analogs, including Remdesivir (16–18), which is a prodrug that is converted to the active drug in the triphosphate form

(RTP) within cells (19). As such, RdRp has been a subject of intensive structural biology efforts. The structures of nsp7, nsp8, and the complex of nsp12-nsp7-nsp8 have been determined (15, 20–23), providing the overall architecture of the RdRp complex. However, the drug discovery effort is hampered because there is no structure of the SARS-CoV-2 RdRp in complex with an RNA template or with nucleotide inhibitors. Here we determined two cryo-EM structures of the SARS-CoV-2 RdRp complex either in the apo form or in a complex with a template-primer RNA and the antiviral drug Remdesivir.

For cryo-EM studies, we co-expressed nsp12 with nsp7 and nsp8 to form the core RdRp complex in insect cells (Fig. 1A and fig. S1, A to D). The stoichiometric ratio of nsp7 and nsp8 appeared to be less than nsp12, thus additional bacterially expressed nsp7 and nsp8 were supplemented before the final purification step to improve the yield of heterotrimeric complex. The purified nsp12 alone showed little activity in binding to a 50-base partial double-stranded template-primer RNA (Fig. 1B and fig. S1E), which is similar to the SARS-CoV nsp12 (14). The presence of nsp7 and nsp8 dramatically increased nsp12 binding to the template-primer RNA (fig. S1E). The nsp12-nsp7-nsp8 complex also showed RNA

polymerization activity on a poly-U template upon addition of adenosine triphosphate (ATP) (Fig. 1, B and C). This RNA polymerization activity was effectively inhibited by the addition of the active triphosphate form of Remdesivir (RTP) (Fig. 1D). Even in the presence of 10 mM ATP, 1 mM RTP completely inhibited RdRp polymerization activity. In contrast, Remdesivir, as a prodrug, at 5 mM concentration, did not have any inhibitory effect on the polymerization activity of the purified enzyme (fig. S1F), neither did Remdesivir in its monophosphate form (RMP) (fig. S1G).

The purified RdRp complex is relatively thermostable with a melting temperature of 53°C (fig. S1H). Negative-stain EM visualization of the nsp12-nsp7-nsp8 complex displayed monodispersed particles with excellent homogeneity (fig. S1I). For the apo nsp12-nsp7-nsp8 complex, we vitrified the sample in the presence of the detergent DDM. The initial attempt at image processing revealed that the particles are preferentially oriented (fig. S2A). Therefore, we collected over 7,400 micrograph movies of more than 5.7 million particle projections to increase the number of projection from the non-preferential orientation. Of these, 81,494 particles were used to yield a density map of 2.8 Å resolution (fig. S2, B and E). Cryo-EM studies of the nsp12-nsp7-nsp8 complex bound with the template-primer RNA and RTP (termed as the template-RTP RdRp complex) faced two challenges (fig. S3). First, most particles were adsorbed to cryo-EM grid bars rather than staying in the vitreous ice. Second, the RNA duplex were dissociated from the template-RTP RdRp complex likely due to conditions of cryo-EM specimen preparation. Eventually, we prepared the cryo-EM specimen of the template-RTP RdRp complex at 15 mg/ml, much higher than normal concentrations used for EM studies of soluble protein complexes. The high concentration of the complex has a mass action effect to stabilize the RNA-protein complex and has excess amount of the complex to escape the absorption of cryo-EM grid bars to enter the vitreous ice (fig. S3). We collected 2,886 micrograph movies, which yielded a 2.5 Å resolution structure using 130,386 particle projections. Because of the relatively high resolution of our structure, the EM map is clear for key structural features across the complex (fig. S4, A to F).

The structure of the apo RdRp complex contains one nsp12, one nsp7 and two nsp8, with an overall arrangement resembling those seen in SARS-CoV and the recently solved structure of SARS-CoV-2 (15, 23) (Fig. 2, A and B). Different from the SARS-CoV RdRp structure but similar to the recent SARS-CoV-2 RdRp structure, our structure reveals that nsp12 also contains an N-terminal  $\beta$ -hairpin (residues 31-50) and an extended nidovirus RdRp-associated nucleotidyl-transferase domain (NiRAN, residues 115-250) (24), with seven helices and three  $\beta$ -strands (15, 23). Following the NiRAN domain is an interface domain (residues 251-365) composed of three helices and five  $\beta$ -strands, which is connected to the RdRp

domain (residues 366-920) (Figs. 1A and 2B). The nsp12 RdRp domain displays the canonical cupped right-handed configuration (25), with the finger subdomain (residues 397-581 and residues 621-679) forming a closed circle with the thumb subdomain (residues 819-920) (Fig. 2, A and B). The closed conformation is stabilized by the binding of nsp7 and nsp8, with one nsp8 molecule sitting on the top of the finger subdomain and interacting with the interface domain. The closed conformation of nsp12 is further stabilized by the nsp7-nsp8 heterodimer, which is packed against the thumb-finger interface (Fig. 2, A and B). In addition, we were able to assign two zinc ions in the conserved metal binding motifs composed by H295-C301-C306-C310 and C487-H642-C645-C646 (Fig. 2C), which are also observed in the SARS-CoV RdRp structure (15). These zinc ions likely serve as conserved structural components in maintaining the integrity of the RdRp architecture.

The structure of the template-RTP RdRp complex contains one nsp12, one nsp7 and one nsp8 (Fig. 3, A and B). The second nsp8 was largely invisible in the EM map of the template-RTP complex (fig. S4C), therefore it was not included in the final model. In addition, the template-RTP RdRp structure contains 14-base RNA in the template strand, 11-base RNA in the primer strand, and the inhibitor Remdesivir in its monophosphate form (RMP) (Fig. 3, C and D), which is covalently linked to the primer strand, as well as a pyrophosphate and three magnesium ions that may serve as catalytic ions near the active site (Fig. 3D and fig. S4, D to F) (26).

The overall structure of the template-RTP RdRp complex is similar to the apo RdRp structure, with nsp12 in a closed conformation (Figs. 2A and 3A). The double-stranded RNA helix, formed by 11 base-pairs from the template-primer RNA (Figs. 3C and 4, A to E), is held by the finger-palm-thumb subdomains. Extensive protein-RNA interactions are observed between the template-primer RNA and nsp12, with a total of 29 residues from nsp12 directly participating in the binding of the RNA (Fig. 4E). Surprisingly no RNA interactions are mediated by nsp7 or nsp8 although these two proteins are required for RNA binding by RdRp. Most protein-RNA interactions involve the RNA phosphate-ribose backbones, with many interactions directly to 2'-OH groups (Fig. 4E), thus providing a basis to distinguish RNA from DNA. There are no contacts from nsp12 to any base pairs of the template-primer RNA, suggesting a sequence independent binding of RNA by RdRp. This is consistent with the fact that no specific sequence is required for the enzymatic activity of RdRp at the elongation step.

At the 3' end of the primer strand is Remdesivir monophosphate (RMP) (Figs. 3D and 4, D and E, and fig. S4, E and F), which is covalently incorporated into the primer strand at the +1 position (Fig. 4E). Additional nucleotides at the +2 and +3 positions of the template strand interact with residues from the back of finger subdomain (Fig. 4, A and B). Despite

the presence of excess RTP in complex assembly, only a single RMP is assembled into the primer strand as observed in the structure. Thus, Remdesivir, like many nucleotide analog prodrugs, inhibits the viral RdRp activity through non-obligate RNA chain termination, a mechanism that requires the conversion of the parent drug to the active triphosphate form (27, 28).

The RMP position is at the center of the catalytic active site (Fig. 3D). As an adenosine monophosphate analog, RMP forms base-stacking interactions with upstream base from the primer strand and two hydrogen bonds with the uridine base from the template strand (Fig. 3D and fig. S5). In addition, RMP also forms interactions with side chains from K545 and R555. Near the bound RMP are two magnesium ions and a pyrophosphate. Both magnesium ions interact with the phosphate diester backbone and they are part of catalytic active site. The pyrophosphate is at the gate of the nucleotide entry channel to the active site and may block the entry of nucleotide triphosphate to the active site (Fig. 3, C and D).

The catalytic active site of the nsp12 RdRp is constructed by seven conserved motifs from A to G (Figs. 1A and 3E and fig. S6). Motifs ABCD are from the palm subdomain with an SDD sequence (residues 759-761) in motif C forming the catalytic active center (Fig. 3D). Both D760 and D761 are involved in coordination of the two magnesium ions at the catalytic center. Motifs F and G are located within the finger subdomain and they interact with the template strand RNA and direct this strand into the active site (Fig. 3E). Motif F also interacts with the primer strand RNA with the side chains of K545 and R555 contacting the +1 base (Fig. 3D), thus stabilizing the incoming nucleotide in the correct position for catalysis. The orientation of template-primer RNA in the active site is similar to the orientation of template-primer RNA in the poliovirus RdRp elongation complex (29) and the HCV NS5B RdRp inhibitor complex (30) (fig. S7). The residues involved in RNA binding as well as residues comprising the catalytic active site are highly conserved (31, 32), highlighting the conserved mechanism of genome replication by RdRp in these diverse RNA viruses, and suggesting it may be possible to develop broad spectrum antiviral inhibitors such as Remdesivir (18) and Galidesivir (BCX4430) (33).

Structural comparison reveals several interesting differences between the apo and complex structures (Fig. 3, E and F, and fig. S8, A and B). First, nsp7 moves toward the RdRp core by 1.5 Å (as measured by nsp7 residue F49, fig. S8, A and B), leading to a rearrangement of the interface, which results in weaker association of the second nsp8 in the complex. Second, the loop that connects the first and second helix of the thumb subdomain moves outward by 2.8 Å (as measured by nsp12 residue I847) to accommodate the binding of the double stranded RNA helix (Fig. 3F). Third, motif G residues K500 and S501 also move outward by 2.0 Å to accommodate

the binding of the template strand RNA. Outside of these changes, the apo nsp12 and the RNA-complex nsp12 are very similar, with an RMSD of 0.52 Å for all C $\alpha$  atoms across the whole protein. In particular, the structural elements that make up the catalytic active site can be exactly superimposed (Fig. 3E), suggesting that the SARS-CoV-2 RdRp is a relatively stable enzyme that is ready to function as a replicase upon binding of RNA template. Viral RdRp is a highly processive enzyme with a rate of replication up to 100 nucleotides per second (34). No significant conformational changes between the apo and the active enzyme structures are consistent with the high processivity of the viral RNA polymerase, which does not need to consume additional energy for conformational changes in the active site during the replication cycle.

Besides Remdesivir, several nucleotide analog drugs, including Favipiravir, Ribavirin, Galidesivir, and EIDD-2801, efficiently inhibit SARS-CoV-2 replication in cell-based assays (35, 36). Like Remdesivir, these nucleotide analogs are proposed to inhibit the viral RdRp through non-obligate RNA chain termination, a mechanism that requires the conversion of the parent compound to the triphosphate active form (33). The structure of the template-RTP RdRp complex provides an excellent model to rationalize how these drugs inhibit the SARS-CoV-2 RdRp activity (fig. S8C). In particular, EIDD-2801 has been shown to be 3-10 times more potent than Remdesivir in blocking SARS-CoV-2 replication (36). The N4 hydroxyl group off the cytidine ring forms an extra hydrogen bond with the side chain of K545 and cytidine base also forms an extra hydrogen bond with the guanine base from the template strand. These two extra hydrogen bonds may explain the apparent higher potency of EIDD-2801 in inhibiting SARS-CoV-2 replication.

The COVID-19 pandemic has inflicted emotional pain and economic burden across the globe. Enzymes that are vital for the viral life cycle are excellent antiviral drug targets as they are different from the host proteins. Among viral enzymes, RdRp is the major target of many existing nucleotide drugs. In this paper, we report the structure of the SARS-CoV-2 RdRp complex in the apo form and in the complex with a template-primer RNA and the active form of Remdesivir. The structures reveal how the template-primer RNA is recognized by the enzyme and how chain elongation is inhibited by Remdesivir. Structure comparison and sequence alignment suggest that the mode of substrate RNA recognition and Remdesivir inhibition of RdRp is highly conserved in diverse RNA viruses, providing a basis for designing broad spectrum antiviral drugs based on nucleotide analogs. Moreover, our structures provide a solid template for modeling and modifying the existing nucleotide drugs, including the highly potent EIDD-2801. Together, these observations provide a rational basis to design even more potent inhibitors to combat the vicious infection of SARS-CoV-2.

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## SUPPLEMENTARY MATERIALS

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Materials and Methods

Figs. S1 to S8

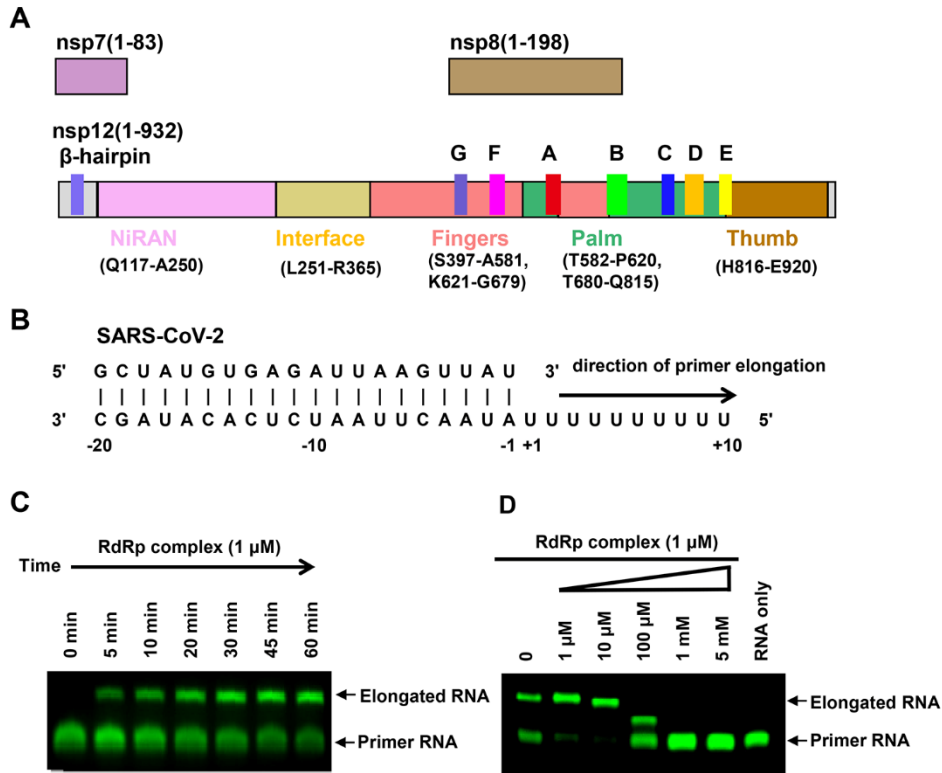
Table S1

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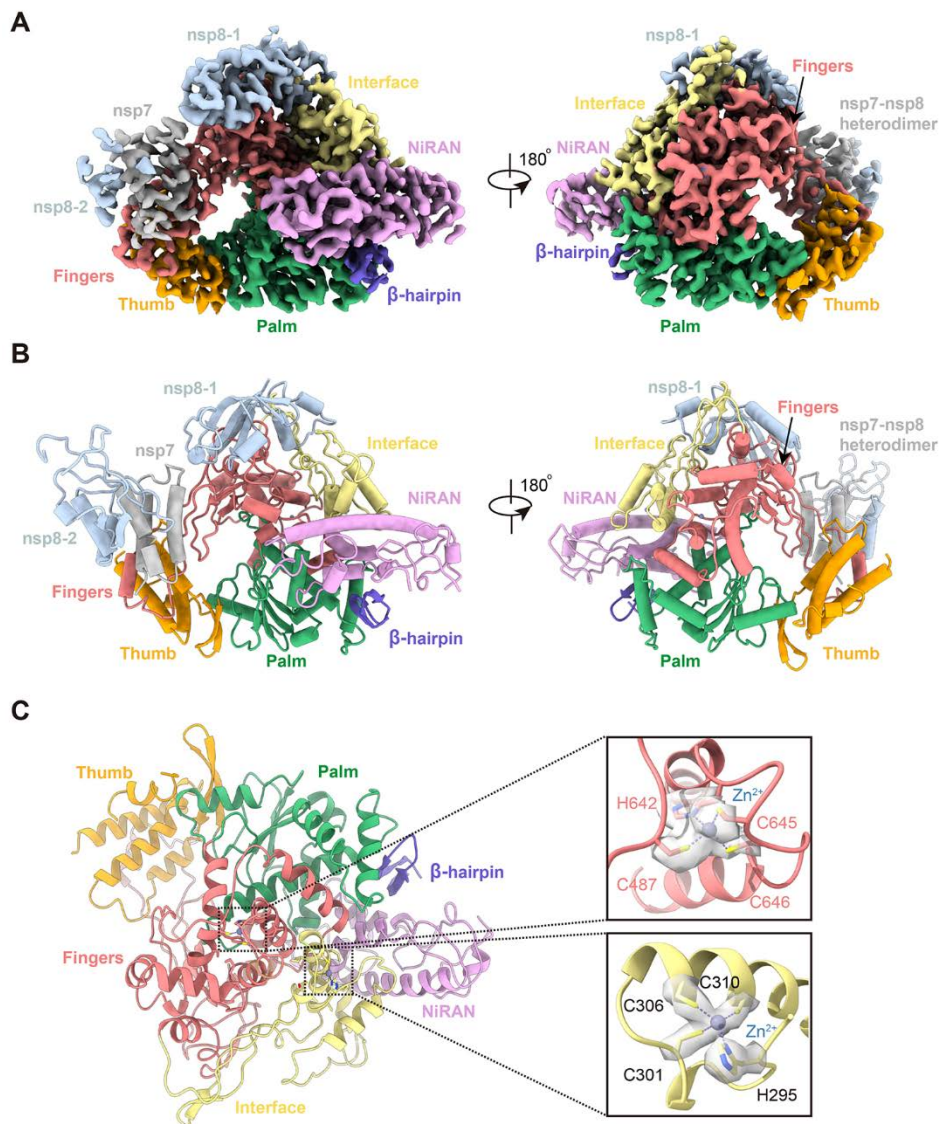
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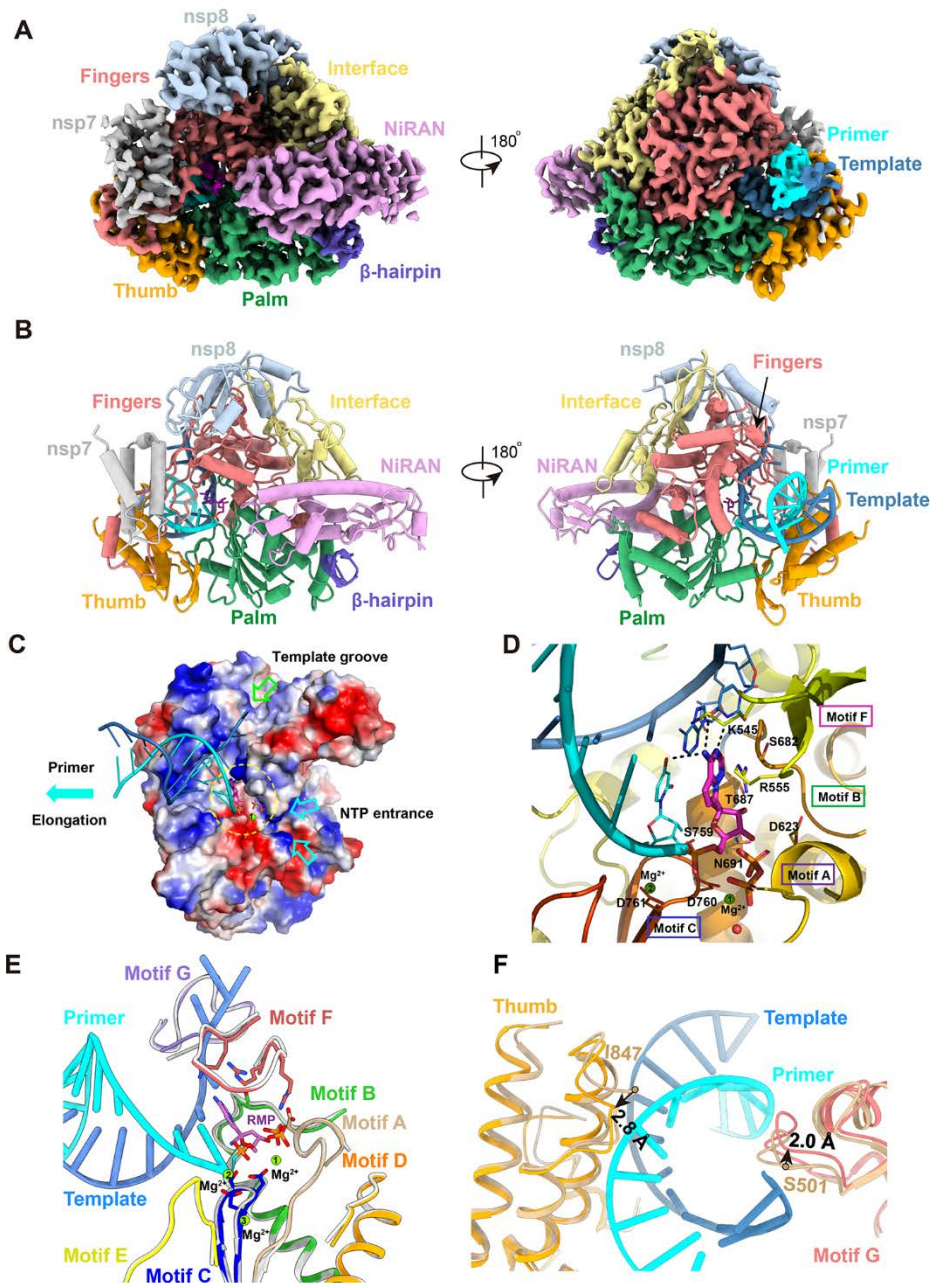
10.1126/science.abc1560



**Fig. 1. Assembly of an active nsp12-nsp7-nsp8 RdRp complex and its inhibition by Remdesivir.** (A) Schematic diagram for the components of the RdRp complex, containing nsp12, nsp7 and nsp8. The polymerase motif (A-G) and  $\beta$ -hairpin unique to the SARS-CoV-2 are highlighted. (B) Sequence of the RNA duplex with a 5'  $U_{10}$  overhang as template for primer extension and RdRp-RNA complex assembly. (C) The recombinant RdRp complex shows polymerase activity in vitro. The primer strand is labeled with fluorescence at the 5' end. (D) Elongation of partial RNA duplex by the purified RdRp complex and its inhibition by RTP.

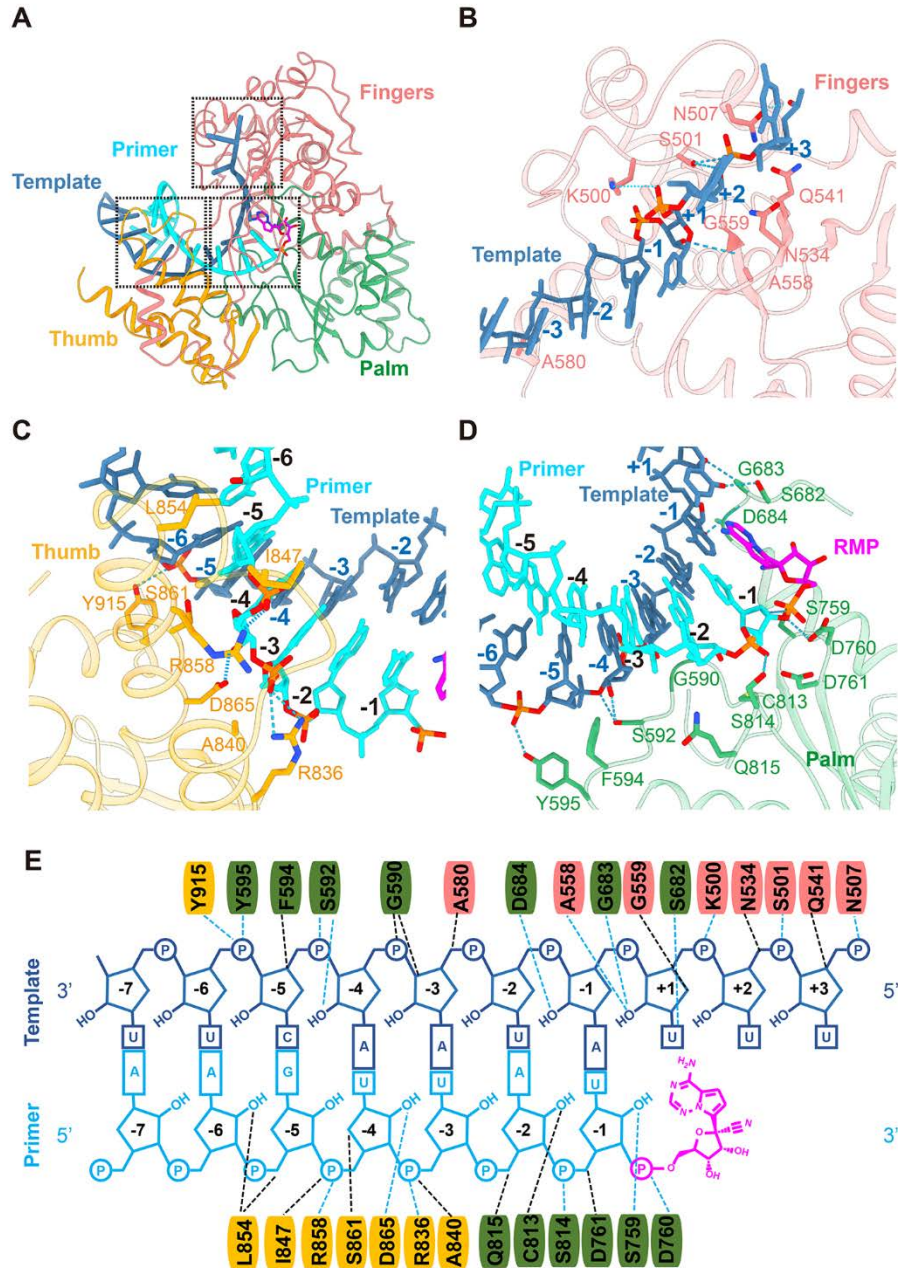


**Fig. 2. Cryo-EM Structure of the apo nsp12-nsp7-nsp8 RdRp complex.** (A and B) Two views of cryo-EM map (A) and structure (B) of the apo nsp12-nsp7-nsp8 complex. The color scheme is according to Fig. 1A and is used throughout the paper. (C) The conserved zinc binding motifs are highlighted in the apo structure rendered in ribbon. The coordinate details of the zinc-binding residues are shown in stick with the EM map in gray surface.



**Fig. 3. Cryo-EM Structure of the Remdesivir and RNA bound RdRp complex.** (A and B) Two views of cryo-EM map (A) and structure (B) of the nsp12-nsp7-nsp8 in complex with template-primer RNA and Remdesivir. (C) Surface view of the RdRp active site with the electrostatic potential from red (negative) to blue (positive). For clarity, residues 410-442, 834-919 of nsp12 and nsp8 are excluded from the figure. The covalently bound Remdesivir in the monophosphate form and the product, pyrophosphate, are shown. The active site is emphasized with a yellow dashed circle. The template groove, the entrance for NTP and the elongation direction are annotated with different colored arrows. (D) A close view of the RdRp active site, showing the covalently bound RMP, pyrophosphate, and magnesium ions. Key residues and bases that interact with Remdesivir are shown. (E and F) Superposition of the conserved RdRp motifs (A to G) of the RNA bound complex with the apo structure colored in gray, with a close view at the active site (E) and at the exit of the template and primer strand (F).





**Fig. 4. RNA Recognition by the RdRp complex.** (A to D) Protein-RNA interactions in the RNA and Remdesivir bound RdRp complex. (E) Diagram of detailed RNA interactions by RdRp.